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Two new sesquiterpenoids from the soft coral Sinularia polydactyla

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Two new sesquiterpenoids from the soft coral Sinularia polydactyla (Ehreberg)

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Two new sesquiterpenoids, polydactins A (1) and B (2) and a known sesquiterpene, 10α -hydroxycadin-4-en-15-al (3), were isolated from the soft coral *Sinularia polydactyla* (Ehreberg). Their structures were determined mainly by spectroscopic methods. Polydactin A (1) showed moderate cytotoxic activities against human oral epidermoid carcinoma cell lines (KB) and human breast carcinoma (MCF) tumour cell lines (*in vitro*).

Keywords: soft coral; Sinularia polydactyla; sesquiterpenoids; anti-tumour; polydactin A; polydactin B

1. Introduction

The corals and marine sponges are abundant natural resources in the South Sea of China. Many bioactive terpenoids have been found from these marine organisms.¹⁻⁷ As a part of our continuing search for bioactive substances from marine organisms, the soft coral Sinularia polydactyla (Ehreberg) was investigated. Bowden et al.⁸ and Duh et al.⁹ have isolated five diterpenes and two norditerpenes from this soft coral, respectively. In our study, two new sesquiterpenes, named polydactins A (1) and B (2), together with a known sesquiterpene, 10α -hydroxycadin-4-en-15-al (3)¹⁰ were isolated but no diterpenes were found from the same soft coral. Bioassay exhibited that 1 has moderate cytotoxic activities against human oral epidermoid carcinoma cell lines (KB) and human breast carcinoma (MCF) tumour cell lines.

2. Results and discussion

Polydactin A (1) was isolated as colourless oil. HREI-MS established the molecular formula $C_{14}H_{22}O_2$ with four degrees of unsaturation. NMR spectral data suggested the presence of an α,β -conjugated keto group [δ_C 203.6 (s), 153.7 (d), 130.3 (d); δ_H 6.34 (1H, dd, J = 16.0, 10.5 Hz), 5.94 (1H, d, J = 16.0 Hz), a secondary hydroxyl carbon [δ_C 75.4 (d); δ_H 3.81 (1H, dd, J = 4.5, 10.0 Hz)], an isopropyl group [δ_C 31.5 (d), 20.6 (q), 20.8 (q); δ_H 1.62 (1H, m), 0.93 (3H, d, J = 6.0 Hz), 0.85 (3H, d, J = 6.0 Hz)] and a terminal double bond [δ_C 151.9 (s), 112.7 (t); δ_H 5.33 (1H, d, J = 2.0 Hz), 5.14

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ISSN 1028-6020 print/ISSN 1477-2213 online © 2008 Taylor & Francis DOI: 10.1080/10286020701782361 http://www.informaworld.com (1H, d, J = 2.0 Hz)]. According to the molecular formula and the functionalities mentioned above, compound 1 was suggested to be monocyclic nor-sesquiterpene.

The gross structure of **1** was determined by a detailed analysis of 1D and 2D NMR spectra. The HMQC experiment led to the assignment of all the protons (Table 1). ${}^{1}\text{H} - {}^{1}\text{H}$ COSY revealed the sequences of the correlations depicted by the bold lines in Figure 2. The three sequences along with the functions mentioned above were assembled by HMBC correlations from H-14 to C-3 and C-5; H-3 to C-4; H-5 to C-4; H-1 to C-10; H-9 to C-10; H-7 to C-8 and H-8 to C-11 permitted the connectivity of the isolated spin systems (Figure 2).

The relative stereochemistry of **1** was established through the coupling constant and NOESY spectrum. An *E*-configuration for the $\Delta^{8,9}$ double bond was determined by the large coupling constant between H-8 and H-9 (J = 16.0 Hz). A NOESY interaction between H-3 ($\delta_{\rm H}$ 3.81)/Hb-5 ($\delta_{\rm H}1.65$), Hb-5/H-7 ($\delta_{\rm H}$ 1.97) and Ha-5 ($\delta_{\rm H}$ 2.64)/H-11 ($\delta_{\rm H}1.62$) (Figure 3) revealed that H-3 and H-7 were located on the same face of the molecule. Thus, the structure of polydactin A (**1**) was determined as shown in Figure 1.

Polydactin B (2) was a pale yellow transparent oil. HREI-MS established the molecular formula $C_{15}H_{26}O_2$ with three degrees of unsaturation. ¹H NMR and ¹³C NMR spectral data suggested the presence of two secondary hydroxyl carbons [δ_C 78.9 (d), 67.0 (d); δ_H 3.41 (1H, dd, J = 5.0, 11.5 Hz), 3.71 (1H, t, J = 10.0 Hz)], an isopropyl group [δ_C 25.9 (d), 21.0 (q), 16.1 (q); δ_H 2.20 (1H, m), 0.93 (3H, d, J = 7.0 Hz); 0.86 (3H, d, J = 7.0 Hz)], and a terminal double bond

Position	1		2	
	¹³ C	1 H (J in Hz)	¹³ C	$^{1}\mathrm{H}$ (J in Hz)
1	36.2 (CH ₂)	2.99 m 2.25 m	78.9 (CH)	3.41 dd (5.0, 11.5)
2	31.4 (CH ₂)	2.05 m	31.8 (CH ₂)	1.58 m 1.87 m
3	75.4 (CH)	3.81 dd (10.0, 4.5)	35.0 (CH ₂)	2.14 m 2.06 m
4	151.9 (C)		146.1 (C)	
5	33.5 (CH ₂)	2.64 m, Ha 1.65 m, Hb	55.7 (CH)	1.74 d (10.0)
6	34.9 (CH ₂)	2.12 m 1.72 m	67.0 (CH)	3.71 t (10.0)
7	53.0 (CH)	1.97 m	49.2 (CH)	1.28 m
8	153.7 (CH)	6.34 dd (16.0, 10.5)	18.0 (CH ₂)	1.50 m 1.26 m
9	130.3 (CH)	5.94 d (16.0)	36.2 (CH ₂)	1.91 m 1.16 dd (3.0, 11.0)
10	203.6 (C)		41.6 (C)	
11	31.5 (CH)	1.62 m	25.9 (C)	2.20 m
12	20.6 (CH ₃)	0.93 d (6.0), 3H	16.1(CH ₃)	0.86 d (7.0), 3H
13	20.8 (CH ₃)	0.85 d (6.0), 3H	21.0 (CH ₃)	0.93 d (7.0), 3H
14	112.7 (CH ₂)	5.33 d (2.0) 5.14 d (2.0)	11.5 (CH ₃)	0.70 s, 3H
15			107.7 (CH ₂)	5.01 s, Ha 4.73 s, Hb

Table 1. ¹H NMR and ¹³C NMR spectral data of compounds 1 and 2 (recorded at 500/125 MHz in CDCl₃; δ in ppm, J in Hz).

 $[\delta_{C} 146.1 \text{ (s)}, 107.7 \text{ (t)}; \delta_{H} 5.01 (1H, s), 4.73 (1H, s)].$ According to the molecular formula and the functionalities mentioned above, compound **2** was suggested to be a bicyclic sesquiterpene.

 ${}^{1}\text{H}{-}{}^{1}\text{H}$ COSY revealed two sequences depicted by the bold lines in Figure 2. The two sequences along with the terminal double bond, methyl group and a quaternary carbon were assembled by HMBC experiment. HMBC correlations between C-4 (δ_{C} 146.1) and H-3, H-5 and H-6; C-10 (δ_{C} 41.6) and H-5, H-1, H-9 and H₃-14; C-14 and H-5, H-9 and H-1 permitted the connectivity of the isolated spin systems (Figure 2).

The stereochemistry of **2** was determined on the base of the NOESY spectrum and coupling constants.

A NOESY interactions between H₃-14 ($\delta_{\rm H}$ 0.70)/H-1 ($\delta_{\rm H}$ 3.41) and H-6 ($\delta_{\rm H}$ 3.71) and H-6/H-11 ($\delta_{\rm H}$ 2.20) (Figure 3) revealed that H-1, H-6, 14-methyl group and 7-isopropyl group were located on the same face of the molecule. In addition, NOESY interaction between H-5 ($\delta_{\rm H}$ 1.74) and H-7 ($\delta_{\rm H}$ 1.28) together with the coupling constant $J_{\rm H-5/H-6} = 10.0$ Hz implied that H-5 and H-7 were located on the opposite side of H-6. Thus, the structure of polydactin B (**2**) was determined as shown in Figure 1.

Polydactins A (1) and B (2) were evaluated for cytotoxic activities against human tumour cell lines KB and MCF. 1 exhibited moderate cytotoxic activity against KB and MCF cell lines with IC_{50} values 13.0 and 14.0 µg /ml,



Figure 1. Structures of compounds 1–3.



Figure 2. ${}^{1}H^{-1}H$ COSY correlations (bold lines) and key HMBC correlations ($H \rightarrow C$) of **1** and **2**.



Figure 3. Key NOE correlations of 1 and 2.

respectively, while **2** exhibited weak cytotoxic activity on KB and MCF cell lines with IC_{50} values 49.4 and 37.8 µg /ml, respectively.

3. Experimental

3.1 General experimental procedures

Optical rotations were measured on a Schmidt & Haensch polaptronic hnqw5 polarimeter. IR spectra were recorded with an EQUINOX55 (Bruker) spectrophotometer. ¹H NMR and ¹³C NMR spectra were recorded with a Varian Unity INOVA spectrometer at 500 MHz respectively, with TMS as internal standard. FAB-MS spectra were obtained with MAT95XP and HR-EI-MS was obtained with MAT95XP (Thermo) Mass Spectrometer. HPLC was conducted on a Perkin–Elmer series 200 using a Diode Array detector 235C and a reversed-phase Symmetry Prep C18 column (7 μ m, 7.8 × 300 mm). Preparative TLC was performed with Silica gel H (200–300 mesh) was used for flash chromatography.

3.2 Biological material

The specimens of *Sinularia polydactyla* (Ehreberg) were collected from the Bay of Sanya, Hainan Island, China, in 1998. Zhi-Can Tang of the Qingdao Institute of Oceanology, Academia Sinica, identified the soft coral species. A voucher specimen (No. 98-SY-17) is preserved in the Research Centre of Organic Natural Products, Sun Yat-Sen University.

3.3 Extraction and isolation

The soft coral S. polydactyla (dry weight 3.05 kg) was extracted with EtOH. The EtOH extract was concentrated and the residue (130 g) was partitioned between EtOAc and H₂O. The EtOAc fraction (63 g) was subjected to column chromatography on silica gel, using petroleumether (PE) and EtOAc mixtures of increasing polarity as eluant to yield 11 fractions (1-11). Fraction 1 (500 mg) eluted with PE/EtOAc (9:1) was further subjected to flash chromatography over silica gel H using PE/EtOAc (6:1) to give compound 1 (15 mg). Fraction 2 (1.5 g) eluted with PE/EtOAc (4:1) was further subjected to flash chromatography over silica gel H using PE/EtOAc (4:1) to give two fractions 2A (261 mg) and 2B (537 mg). Fraction 2B was separated by RP-HPLC on C18 silica gel using MeOH/H₂O (17:3) as eluant to yield five fractions $(2B_1 - 2B_5)$. After evaporation of fraction $2B_3$, pure compound 2 (30 mg) was obtained. Fraction $2B_5$ (50 mg) was further separated by RP-HPLC on C18 silica gel using MeOH/H₂O (17:3) as eluant to afford compound 3 (20 mg).

3.3.1 Polydactin A (1)

Colourless oil, $[\alpha]_D^{20} = -28.95$ (*c* 0.076, CHCl₃); IR ν_{max} KBr cm⁻¹: 3431 (OH), 3077 (=C-H), 1717(C=O), 1673, 1461, 1386, 1256, 1037; ¹H NMR and ¹³C NMR spectral data: see table 1. HREI-MS *m/z* 222.1609 [M]⁺ (calcd for C₁₄H₂₂O₂, 222.1614); FAB-MS *m/z* 223 [M + H]⁺.

3.3.2 Polydactin B (2)

Pale yellow oil, $[\alpha]_D^{20} = +23.61$ (*c* 0.072, CHCl₃); IR ν_{max} KBr cm⁻¹: 3418 (OH), 3078 (=C-H), 1706, 1648, 1460, 1383, 1060, 1005; ¹H NMR and ¹³C NMR spectral data: see table 1. HREI-MS *m/z* 238.1894 [M]⁺ (calcd for C₁₅H₂₆O₂, 238.1927); FAB-MS *m/z* 239 [M + H]⁺.

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